

Isolation of drug target protein (2) Protein kinase inhibitors [Bisindolylmaleimide]

Summary

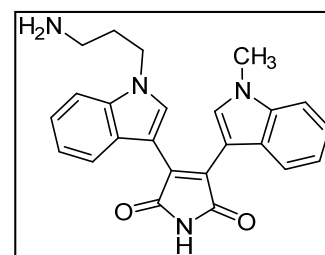
A protein kinase is an enzyme that modifies other proteins by adding phosphate groups to them (phosphorylation).

We show the result of experiment here to purify the targets of Bisindolylmaleimide VIII (Bis VIII) in certain condition. Bisindolylmaleimide compounds were originally reported as a potent inhibitor against Protein Kinase C. The inhibition of the other kinases such as GSK3 α/β ¹⁾, CDK2²⁾ and Rsk³⁾ were also observed afterwards.

HeLa cell extracts we used was not treated by TPA or any other stimulator. And Bis VIII was immobilized on FG beads (NHS beads) to perform affinity purification of bound proteins with Bis VIII immobilized beads.

As the result, several binding proteins were separated and one of major proteins, bound to Bis VIII immobilized beads was GSK3 α/β , confirmed by western blotting and mass spectrometry in this experimental condition.

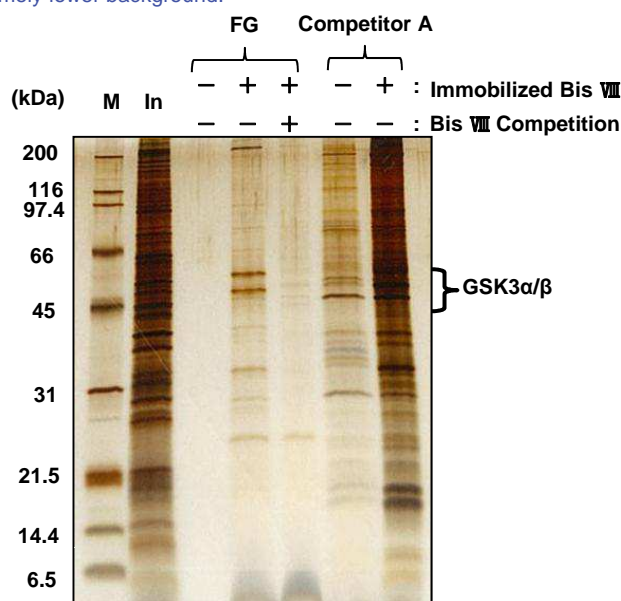
The major purified proteins may be changed dependent on the amount of compounds immobilized on FG beads.



Bisindolylmaleimide VIII

Result

- Several binding proteins were separated using FG beads or Competitor A. When free Bis VIII was added to HeLa cell extracts before incubation with the beads, the yields of several proteins were reduced, which suggested that these proteins specifically interact with Bis VIII. Two major protein bands, which are supposed to be GSK3 α/β ²⁾ from their molecular weight, were selected for further identification experiments. In addition, Their bands were confirmed as GSK3 α/β by mass spectrometry.
- We also compared FG beads with other commercially available magnetic beads. We confirmed FG beads have extremely lower background.



1) Nihar R, Jena. Journal of Molecular Modeling 18 (2012) 631-644

2) Brehmer, D., Godl, K., Zech, B., Wissing, J., and Daub, H. Molecular & Cellular Proteomics 3.5 (2004) 490-500

3) Roberts, NA., Haworth, RS., Avkiran, M., J Pharmacol. 145(4) (2005) 477-489

Materials and method

Materials

1. Bisindolylmaleimide VIII
2. NHS beads
3. HeLa cell extracts (cytosolic fraction) – 3mg/ml
4. Binding & Washing Buffer (20mM HEPES-NaOH(pH7.9), 100mM KCl, 1mM MgCl₂, 0.2mM CaCl₂, 0.2mM EDTA, 10%(v/v) glycerol, 0.1% NP-40, 1mM DTT, 0.2mM PMSF)
5. Elution Buffer (0.0625M Tris-HCl (pH6.8), 0.005% BPB, 2% SDS, 10% glycerol, 5% 2-mercapto ethanol)
6. Mouse monoclonal anti-GSK3α/β (from Santa Cruz Biotechnology)
7. Anti-Mouse IgG, HRP-Linked Whole Ab Sheep (from GE Healthcare)
8. Starting Block™ (TBS) Blocking Buffer (from Thermo scientific)
9. TBS-T buffer (137mM NaCl, 2.68mM KCl, 25mM Tris-HCl, pH7.4, 0.1w/v% Tween-20)

Methods 1 (Immobilization)

1. Apply

FG beads

- 0.5mg NHS beads and 0.1mM Bis VIII in 100ul DMF
- 0.5mg NHS beads and NO Bis VIII in 100ul DMF (Negative Control)

Competitor A

- 0.5mg NHS beads and 1mM Bis VIII in 100ul DMF
- 0.5mg NHS beads and NO Bis VIII in 100ul DMF (Negative Control)

2. Reaction

1) Immobilization

70min at r.t.

2) Masking

Inactivate unreacted NHS according to recommended conditions of each beads.

Methods 2 (Affinity purification)

1. Wash

Wash beads with washing buffer 3 times at 4°C (on ice).

2. Add sample solution

Add 1000ul HeLa cell extracts (Bis VIII Competition : -) or Bis VIII added HeLa cell extracts (Bis VIII Competition : +) to beads. (0.1mM Bis VIII is added to HeLa cell extracts and preincubated for 120min before incubation with the beads.)

3. Reaction

Incubate for 240min at 4°C.

4. Wash

Remove sample solution.
Wash beads with washing buffer 3 times at 4°C (on ice).

5. Elution

Add 40ul elution buffer and resuspend beads.
Boil for 5min at 98°C and remove beads.

6. Analyze the samples by SDS-PAGE and silver staining

Methods 3 (Western blotting)

1. Perform SDS-PAGE and transfer the protein to a PVDF membrane.
2. Block the membrane with the blocking buffer for 15min at room temperature.
3. Dilute the anti-GSK3α/β antibody with the blocking buffer to 1/200.
4. Incubate for 60min at room temperature.
5. Dilute the secondary antibody with the TBS-T buffer to 1/1000.
6. Incubate for 60min at room temperature.
7. Wash the membrane with TBS-T buffer three times.
8. Detect with a chemiluminescence substrate.

FG beads information

Product name	NHS beads
Product number	TAS8848N1141
Storage temperature	-20°C
Storage buffer	Isopropyl alcohol
Size of beads	190nm ± 20nm
Functional groups	200 - 300nmol/mg

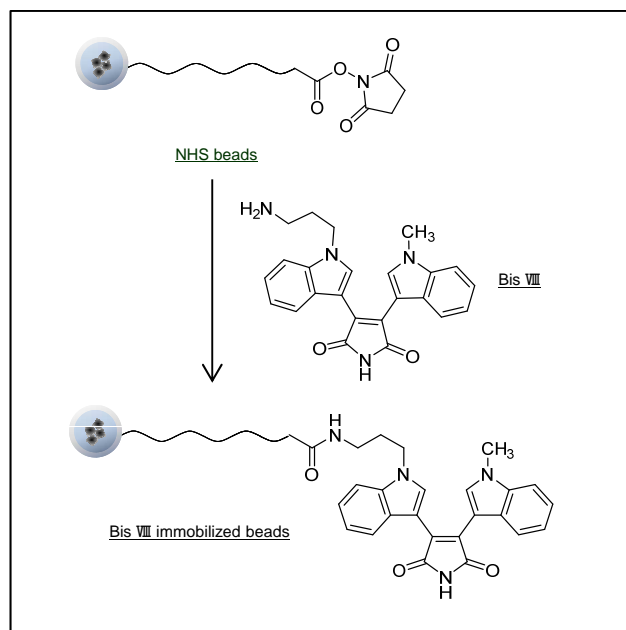


Fig.1 Immobilization of Bis VIII on NHS beads

Methods 4 (MS)

1. Excising the bands from the gel.
2. Wash the band pieces, then dry the band pieces.
3. Add 500ul/band of 10mM DTT/25mM NH₄HCO₃.
4. Incubate for 60min at 56°C.
5. Add 500ul/band of 55mM iodeacetamide/25mM NH₄HCO₃.
6. Incubate for 60min at room temperature.
7. Wash the band pieces with 200ul of 25mM NH₄HCO₃ for 10min.
8. Wash the band pieces with 500ul of 25mM CH₃CN/NH₄HCO₃ (50:50 v/v) for 10min.
9. Dry the band pieces.
10. Add 10ul/band of 10ug/ml trypsin in 25mM NH₄HCO₃.
11. Incubate overnight at 37°C.
12. Add 50ul/band of 25mM CH₃CN/trifluoroacetic acid/ultrapure water (50:5:45 v/v).
13. Incubate for 30min at room temperature.
14. Transfer supernatant to a new clean tube.
15. Dry the peptides in the tube.
16. Injection on the MS.